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# **Conformational studies on peptides containing a,a-disubstituted a-amino acids: chiral cyclic a,a-disubstituted a-amino acid as an a-helical inducer†**

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Four types of  $\alpha$ , $\alpha$ -disubstituted amino acids {*i.e.*,  $\alpha$ -aminoisobutyric acid (Aib), 1-aminocyclopentanecarboxylic acid (Ac5c), (3*S*,4*S*)-1-amino-(3,4-dimethoxy)cyclopentanecarboxylic acid [(*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup>] and its enantiomer (*R*,*R*)-Ac<sub>5</sub>c<sup>dOM</sup>} were introduced into L-leucine-based hexapeptides and nonapeptides. The dominant conformations of eight peptides: Cbz-(L-Leu-L-Leu-dAA)<sub>2</sub>-OMe [dAA = 1; Aib; 2; Ac<sub>5</sub>c; 3; (*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup>; 4: (*R,R*)-Ac<sub>5</sub>c<sup>dOM</sup>] and Boc-(L-Leu-L-Leu-dAA)<sub>3</sub>-OMe [dAA = **5**: Aib; **6**: Ac<sub>5</sub>c; **7**: (*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup>; **8**: (*R*,*R*)-Ac<sub>5</sub>c<sup>dOM</sup>], were investigated by IR, CD spectra and X-ray crystallographic analysis. The CD spectra revealed that Aib hexapeptide **1** and Ac<sub>5</sub>c hexapeptide **2** formed right-handed (*P*)  $3_{10}$ -helices, while Ac<sub>5</sub>c<sup>dOM</sup> hexapeptides **3** and **4** formed a mixture of  $(P)$  3<sub>10</sub>- and  $\alpha$ -helices. The Aib nonapeptide **5** formed a  $(P)$  3<sub>10</sub>-helix, the Ac<sub>5</sub>c nonapeptide 6 formed a mixture of (*P*)  $3_{10}$ - and  $\alpha$ -helices, and the Ac<sub>5</sub>c<sup>dOM</sup> nonapeptides 7 and 8 formed (*P*) a-helices. X-Ray crystallographic analysis revealed that the Aib hexapeptide **1** formed a (*P*)  $3_{10}$ -helix, while (*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup> hexapeptide **3** formed a (*P*)  $\alpha$ -helix. In addition, the Ac<sub>5</sub>c nonapeptide **6** and  $(R,R)$ -Ac<sub>5</sub>c<sup>dOM</sup> nonapeptide **8** formed (*P*)  $\alpha$ -helices. The Aib and achiral Ac<sub>5</sub>c residues have the propensity to form  $3_{10}$ -helices in short peptides, whereas the chiral Ac<sub>5</sub>c<sup>dOM</sup> residues have a penchant for forming  $\alpha$ -helices.

## **Introduction**

Precise control of the helical structures of peptides and proteins is highly important because they play a significant role in a variety of fields such as biological sciences, and material and medicinal chemistries.**1–4** To create such helical structures, utilization of  $\alpha$ , $\alpha$ -disubstituted  $\alpha$ -amino acids (dAAs) is a powerful method because the dAAs could restrict the conformational freedom of their peptides and stabilize the secondary structures.**5–12** Among them,  $\alpha$ -aminoisobutyric acid (Aib), in which the  $\alpha$ -hydrogen atom of L-alanine is replaced with a methyl substituent, has been widely used to stabilize helical structures of peptides.**13–19** We have recently reported that the introduction of Aib residues into the L-leucine-based hexapeptide (L-Leu-L-Leu-Aib-L-Leu-L-Leu-Aib) stabilized a right-handed  $(P)$  3<sub>10</sub>-helix, whereas that of

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 $1$ -amino-(3,4-dimethoxy)cyclopentanecarboxylic acid (Ac<sub>5</sub>c<sup>dOM</sup>) residues,**20,21** which is a chiral cyclic dAA bearing only sidechain chiral centers,<sup>22,23</sup> stabilized a (*P*)  $\alpha$ -helix in the solid state.**24,25** As a part of our ongoing research, we investigated the preferred conformations of four L-leucine-based hexapeptides,  $Cbz$ -(L-Leu-L-Leu-dAA)<sub>2</sub>-OMe [dAA = 1; Aib; 2: Ac<sub>5</sub>c; 3: (*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup>; 4: (*R,R*)-Ac<sub>5</sub>c<sup>dOM</sup>] (Cbz: benzyloxycarbonyl, Ac<sub>5</sub>c: 1aminocyclopentanecarboxylic acid, OMe: methyl ester), and four nonapeptides, Boc-(L-Leu-L-Leu-dAA)<sub>3</sub>-OMe  $\left[ dAA = 5: Aib; 6: \right]$ Ac<sub>5</sub>c; 7: (*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup>; 8: (*R*,*R*)-Ac<sub>5</sub>c<sup>dOM</sup>], in solution and in the crystalline state (Fig. 1). The IR and CD spectra, and X-ray crystallographic analysis revealed that the Aib and achiral  $Ac<sub>5</sub>c$ residues have the tendency to form  $(P)$  3<sub>10</sub>-helices, whereas chiral cyclic dAA:  $(S, S)$ - and  $(R, R)$ -Ac<sub>5</sub>c<sup>dOM</sup>, have a penchant for forming ( $P$ )  $\alpha$ -helices in short L-leucine-based peptides.

#### **Results and discussion**

Cyclic amino acids  $(S, S)$ -Ac<sub>5</sub>c<sup>dOM</sup> and  $(R, R)$ -Ac<sub>5</sub>c<sup>dOM</sup> were synthesized according to previously reported methods.**20,24** The preparation of hexapeptides **1–4** and nonapeptides **5–8** was carried out by a solution-phase method using 1- (3-dimethylaminopropyl)-3-ethylcarbodiimide (EDC) hydrochloride and 1-hydroxybenzotriazole (HOBt) hydrate as coupling reagents.**24–27**

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**Fig. 1** Achiral and chiral  $\alpha$ , $\alpha$ -disubstituted  $\alpha$ -amino acids discussed in this paper.

Fig. 2 shows the FT-IR spectra of **1–8** in the NH-stretching region (amide A 3200–3500 cm<sup>-1</sup>, peptide concentration 1.0 mM in CDCl<sub>3</sub> solution). The weak bands around the  $3430 \text{ cm}^{-1}$  region were assigned to free (solvated) peptide NH groups, and the strong bands around the  $3320 \text{ cm}^{-1}$  region were assigned to peptide NH groups with  $N$ —H $\cdots$  O  $=$  C intramolecular H-bonds. The IR spectra of hexa- and nonapeptides **1–8** were very similar to those of helical peptides in solution,**20,28** but different from those of peptides that form the extended planar  $C_5$  conformation.<sup>29,30</sup> Furthermore, the *Abs*3320/*Abs*<sup>3430</sup> values of nonapeptides **5–8** increased compared with those of hexapeptides **1–4**, indicating that nonapeptides were more helical than hexapeptides.



Fig. 2 FT-IR spectra (3200–3500 cm<sup>-1</sup> region) of (A) hexapeptides Cbz-(L-Leu-L-Leu-dAA)<sub>2</sub>-OMe [dAA = 1: Aib; 2: Ac<sub>5</sub>c; 3: (*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup>; 4:  $(R, R)$ -Ac<sub>5</sub>c<sup>dOM</sup>] and (B) nonapeptides Boc-(L-Leu-L-Leu-dAA)<sub>3</sub>-OMe  $[dAA = 5: Aib; 6: Ac<sub>5</sub>c; 7: (S,S)-Ac<sub>5</sub>c<sup>dOM</sup>; 8: (R,R)-Ac<sub>5</sub>c<sup>dOM</sup>] in CDCl<sub>3</sub>$ solution. Peptide concentration: 1.0 mM.

The CD spectra of peptides **1–8** were measured in a mixture of 2,2,2-trifluoroethanol (TFE) and  $H_2O$  (50/50) solution, and also in the solid state (KCl disk) to obtain information about their secondary structures. All CD spectra of **1–8** showed negative maxima at around 208 and 222 nm, indicating that the screw sense of the helices was right-handed (*P*) in solution and in the solid state (Fig. 3; red-shift of the maximum at 222 nm was observed in the solid state).<sup>24,25</sup> The ratio of *R* ( $\theta_{222}/\theta_{208}$ ) in solution suggested that the dominant secondary structure of Aib hexapeptide **1** and Ac<sub>5</sub>c hexapeptide 2 was a  $3_{10}$ -helix ( $R = 0.5$ ), and that of Ac<sub>5</sub>c<sup>dOM</sup> hexapeptides **3** and **4** ( $R = 0.6$ ) was a mixture of 3<sub>10</sub>- and  $\alpha$ -helices (Fig. 3A).**24,31,32** In the solid state, the *R* values of **1** and **2** were 0.5 and those of **3** and **4** were 1.0 (Fig. 3B). These *R* values indicate that **1** with Aib and **2** with Ac<sub>5</sub>c formed mainly (*P*)  $3_{10}$ -helices, while **3** and **4** with Ac<sub>5</sub>c<sup>dOM</sup> formed  $\alpha$ -helices. In the case of elongated nonapeptides, the preferred conformation of Aib nonapeptide **5** in solution was a  $3_{10}$ -helix ( $R = 0.4$ ), that of achiral Ac<sub>5</sub>c nonapeptide  $6(R=0.6)$  was a mixture of 3<sub>10</sub>- and  $\alpha$ -helices, and those of Ac<sub>5</sub>c<sup>dOM</sup> nonamers **7** and **8** ( $R = > 0.7$ ) were  $\alpha$ -helices (Fig. 3C). In the solid



**Fig. 3** CD spectra in the 190–260 nm region of peptides **1–8**. (A) Hexapeptides Cbz-(L-Leu-L-Leu-dAA)<sub>2</sub>-OMe  $[dAA = 1$ : Aib; 2: Ac<sub>5</sub>c; 3:  $(S, S)$ -Ac<sub>5</sub>c<sup>dOM</sup>; **4**:  $(R, R)$ -Ac<sub>5</sub>c<sup>dOM</sup>] in TFE/H<sub>2</sub>O = 50/50 solution, and (B) in KCl disk. (C) Nonapeptides Boc-(L-Leu-L-Leu-dAA)<sub>3</sub>-OMe [dAA = **5**: Aib; **6**: Ac<sub>5</sub>c; **7**: (*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup>; **8**: (*R*,*R*)-Ac<sub>5</sub>c<sup>dOM</sup>] in TFE/H<sub>2</sub>O = 50/50 solution, and (D) in KCl disk. Peptide concentration: 0.5 mM in TFE/H<sub>2</sub>O solution; 0.5 mg peptides/70 mg KCl in the solid state.

state, a  $(P)$  3<sub>10</sub>-helix seemed to be present in 5 and 6 ( $R = 0.5$ ), and  $\alpha$ -helices seemed to be present in 7 and 8 ( $R = 1.1$ ) (Fig. 3D).

The hexapeptides **1** and **3** and nonapeptides **6** and **8** became suitable crystals for X-ray crystallographic analysis by slow evaporation of the solvent (MeOH/H2O for **1**, **3** and **8**, and EtOH/H<sub>2</sub>O for 6) at room temperature. The crystal and diffraction parameters of **1**, **3**, **6** and **8** are summarized in Table 1, and relevant backbone and side-chain torsion angles and the intra- and intermolecular hydrogen bond parameters are listed in Tables 2 and 3, respectively.

The crystal structure of Cbz-(L-Leu-L-Leu-Aib)<sub>2</sub>-OMe (1) was a right-handed  $(P)$  3<sub>10</sub>-helix (Fig. 4) and solved in the space group *P*1. The mean values of the  $\phi$  and  $\psi$  torsion angles of the amino acid residues (1–5) were -66.1*◦* and -31.3*◦*, respectively, which are close to those for an ideal right-handed  $(P)$  3<sub>10</sub>-helical structure (-60*◦* and -30*◦*).**<sup>33</sup>** Reversal of the torsion angle signs at the C-terminus occurred; that is, the signs of the  $\phi$  and  $\psi$ torsion angles (+50.0*◦* and +39.0*◦*) of the Aib(6) residue were positive. Three consecutive intramolecular hydrogen bonds of the  $i \leftarrow i+3$  type were present between H–N(4) and C(1)=O(1)





**Table 2** Selected torsion angles *w*, *f*, *y* and *c* ( *◦*) for hexapeptides **1** and **3** and nonapeptides **6** and **8** as determined by X-ray crystallographic analysis







#### **Table 3** *(Contd.)*



<sup>*a*</sup> The amino acid numbering begins at the N-terminus of the peptide chain. <sup>*b*</sup> The hydrogen bond between N<sub>4</sub>–H and  $O_0$  (3.71 Å) in hexapeptide 1 is not observed. *c* The D  $\cdots$  A distance is too long for a hydrogen bond. *d* The D  $\cdots$  A distance is a bit long for a hydrogen bond. *e* O<sub>w</sub>: water; O<sub>MeOH</sub>: MeOH; O<sub>M</sub>: methoxy group. *f* The hydrogen bond between  $N_{4b}$ –H and  $O_{0b}$  (3.76 Å) in hexapeptide 3 is not observed.



Fig. 4 X-Ray diffraction structure of Cbz-(L-Leu-L-Leu-Aib)<sub>2</sub>-OMe (1) as viewed (A) perpendicular to and (B) along the helical axis.

 $[N(4)\cdots O(1) = 3.20 \text{ Å}; N-H \cdots O(146.2^{\circ}), H-N(5) \text{ and } C(2) = O(2)$  $[N(5)\cdots O(2) = 2.99 \text{ Å}; N-H\cdots O 159.5^{\circ}],$  and H–N(6) and  $C(3) = O(3)$  [N(6)  $\cdots$  O(3) = 3.19 Å; N–H $\cdots$  O 149.4<sup>°</sup>], although the distances between H–N(3) and C(0)=O(0) [N(3) $\cdots$ O(0) = 3.42 Å; N–H $\cdots$ O 123.2<sup>°</sup>] ( $\beta$ -turn) and H–N(4) and C(0)=O(0)  $[N(4) \cdots O(0) = 3.71$  Å; N–H $\cdots$  O 140.8<sup>°</sup> (α-turn) were too long for hydrogen bonds.

In the packing mode, two intermolecular hydrogen bonds were observed between the  $3_{10}$ -helical conformers; that is, between the H–N(1) donor and the  $C(4')=O(4')$  O atom of a symmetry-related molecule (x, y,  $-1+z$ ) [N(1)  $\cdots$  O(4') = 2.92 Å; N–H $\cdots$ O 164.1<sup>°</sup>] and between the H–N(2) donor and the  $C(5')=O(5')$  O atom of a symmetry-related molecule (x, y,  $-1+z$ ) [N(2) $\cdots$  O(5') = 2.87 Å; N–H···O 166.7°]. The helical molecules were connected by intermolecular hydrogen bonds forming a head-to-tail alignment of chains (Fig. 5).



**Fig. 5** Packing of **1** in the crystalline state. Intermolecular hydrogen bonds are indicated as red dashed lines.

Cbz-[L-Leu-L-Leu- $(S, S)$ -Ac<sub>5</sub>c<sup>dOM</sup>]<sub>2</sub>-OMe (3) exclusively crystallized into two right-handed  $(P)$   $\alpha$ -helical conformers **A** and **B** with a methanol and a water molecule (Fig. 6). The peptide-main chain conformations of molecules **A** and **B** were similar except



**Fig. 6** (A) X-Ray diffraction structure of Cbz-[L-Leu-L-Leu- $(S.S)$ -Ac $s$ c<sup>dOM</sup>]<sub>2</sub>-OMe (3). The methanol and water molecules are omitted. The structure of molecule **A**, as viewed (B) perpendicular to and (C) along the helical axis.



**Fig. 7** The overlaid structures of molecules **A** (green) and **B** (blue), as viewed (A) perpendicular to and (B) along the helical axis.

for differences in the conformations of the N-terminal protecting group and their side chains, especially at  $L-Leu(5)$  and  $Ac_5c^{dOM}(6)$ residues, as shown by their superimposition in Fig. 7. The mean values of the  $\phi$  and  $\psi$  torsion angles of the amino acid residues (1–5) were -63.7*◦* and -40.4*◦* for molecule **A** and -75.8*◦* and -28.4*◦* for molecule **B**, and reversal of the torsion angle signs at the C-terminus occurred. The average torsion angles of molecule **A** were close to those for an ideal right-handed (*P*)  $\alpha$ -helical structure (-60*◦* and -45*◦*), while those of molecule **B** seemed to be a distorted  $(P)$   $\alpha$ -helix with semi-extended conformation at L-Leu(5B) ( $\phi = -99.6^\circ$ ,  $\psi = -11.4^\circ$ ) and Ac<sub>5</sub>c<sup>dOM</sup>(6B) ( $\phi =$ +64.9*◦*, *y* = -167.7*◦*). In molecule **A**, two intramolecular hydrogen bonds of the  $i \leftarrow i+4$  type were observed between H–N(5a) and C(1a)=O(1a)  $[N(5a)\cdots O(1a) = 2.99 \text{ Å}; N-H\cdots O$  150.4<sup>°</sup> and H–N(6a) and C(2a) = O(2a) [N(6a)  $\cdots$  O(2a) = 3.03 Å; N– H ··· O 151.1<sup>°</sup>]. Furthermore, two weak intramolecular hydrogen bonds of the  $i \leftarrow i+3$  ( $\beta$ -turn) type were observed between H– N(3a) and C(0a)=O(0a) [N(3a)  $\cdots$  O(0a) = 3.24 Å; N–H $\cdots$ O 125.8°] and between H–N(4a) and C(1a)= $O(1a)$ [N(4a) $\cdots$ O(1a)= 3.27 Å; N–H $\cdots$ O 119.3<sup>°</sup>], but the distance between H–N(4a) and C(0a)=O(0a)  $[N(4a)\cdots O(0a) = 3.49 \text{ Å}; N-H\cdots O$  166.1<sup>°</sup> was too long for a hydrogen bond of the  $i \leftarrow i+4$  ( $\alpha$ -turn) type. Molecule **B** similarly contained two intramolecular hydrogen bonds between H–N(5b) and C(1b)=O(1b)  $[N(5b)\cdots O(1b) =$ 2.92 Å; N-H ⋅ ⋅ O 145.2<sup>°</sup>] and H-N(6b) and C(2b)=O(2b)  $[N(6b)\cdots O(2b) = 2.98 \text{ Å}; N-H\cdots O(136.6^{\circ}), \text{ and the distance}$ 

between H–N(4b) and C(0b)= $O(0b)$  for  $\alpha$ -turn [N(4b) $\cdots$  O(0b) = 3.76 Å; N–H $\cdots$ O 140.1<sup>°</sup>] was too long for a hydrogen bond. Furthermore, one weak intramolecular hydrogen bond of the  $i \leftarrow i+3$  ( $\beta$ -turn) type was observed between H–N(3b) and  $C(0b) = O(0b)$  [N(3b)  $\cdots$  O(0b) = 3.20 Å; N–H $\cdots$  O 132.9<sup>°</sup>]. In the packing mode, six intermolecular hydrogen bonds were observed between the H–N(1b) donor and the  $C(5a) = O(5a)$  acceptor  $[N(1b)\cdots O(5a) = 2.91 \text{ Å}; N-H\cdots O(5a) = 151.1^\circ$ , H–N(2b) and  $C(4a) = O(4a) [N(2b) \cdots O(4a) = 2.94 \text{ Å}; N-H \cdots O(146.6^{\circ})]$ , the H– O<sub>MeOH</sub> of the methanol donor and C(3b)= $O(3b)$  [O<sub>MeOH</sub>  $\cdots$  O(3b) = 2.79 Å; O–H $\cdots$ O 176.9<sup>°</sup>], the H–O<sub>w</sub> of the water donor and O<sub>MeOH</sub> of the methanol acceptor  $[O_W \cdots O_{MeOH} = 2.80 \text{ Å}; O-H \cdots O$ 173.7°], the H–N(2a) and  $O_{w'}$  [N(2a) $\cdots$  $O_{w'}$  = 2.88 Å; N–H $\cdots$ O 156.3*◦*] of a symmetry-related molecule (x, y, 1+z), and the H–Ow and C(5b')= $O(5b')$  [O<sub>w</sub>  $\cdots$  O(5b') = 2.72 Å; N–H $\cdots$ O 173.6<sup>°</sup>] of a symmetry-related molecule (1+x, y, z). Molecules **A** and **B** were alternately connected, as shown in Fig. 8.



**Fig. 8** Packing of **3** (molecule **A**: green, molecule**B**: blue) in the crystalline state. Intermolecular hydrogen bonds are indicated as red dashed lines.

The structure of Boc-(L-Leu-L-Leu-Ac<sub>5</sub>c)<sub>3</sub>-OMe (6) was solved in the space group *P*21. **<sup>34</sup>** Four crystallographically independent molecules **A**, **B**, **C** and **D** were found in the asymmetric unit together with an ethanol and water molecules. All molecules **A–D** were folded into  $\alpha$ -helical structures (Fig. 9), and the main-chain conformations of peptides **A–D** were well matched except for small differences in the conformations of their side chains (Fig. 10). The mean values of the  $\phi$  and  $\psi$  torsion angles of the amino acid residues (1–8) were -66.9*◦* and -40.6*◦* for **A**, -67.5*◦* and -40.7*◦* for **B**, -67.0*◦* and -40.3*◦* for **C**, -68.0*◦* and -40.3*◦* for **D**. Flip of the torsion angles at the C-terminus residue occurred, that is, the values of the  $\phi$  and  $\psi$  torsion angles of the Ac<sub>5</sub>c(9) were +60.0<sup>°</sup>, -153.8*◦* for **A**; +49.2*◦*, +39.4*◦* for **B**; +61.6*◦*, -162.4*◦* for **C**; and +43.1*◦*, -179.1*◦* for **D**. In molecules **A–D**, five hydrogen bonds of the  $i \leftarrow i+4$  type were observed between H–N(4a–d) and C(0a– d)  $=$  O(0a–d) [N(4a–d)  $\cdots$  O(0a–d) = 3.12 Å (a), 3.08 Å (b), 3.15 Å (c), 3.06 A˚ (d); N–H ◊◊◊ O 157.9*◦* (a), 158.3*◦* (b), 159.7*◦* (c), 155.8*◦*



Fig. 9 X-Ray diffraction structure of Boc-(L-Leu-L-Leu-Ac<sub>5</sub>c)<sub>3</sub>-OMe (6), as viewed (A) perpendicular to and (B) along the helical axis. The ethanol and water molecules have been omitted.



**Fig. 10** The overlaid structures of molecules **A** (blue), **B** (purple), **C** (green) and **D** (yellow), as viewed (A) perpendicular to and (B) along the helical axis.

(d)], H–N(5a–d) and C(1a–d)= $O(1a-d)$  [N(5a–d) $\cdots$ O(1a–d) = 2.89 Å (a), 2.86 Å (b), 2.84 Å (c), 2.86 Å (d); N–H  $\cdots$  O 159.3<sup>°</sup> (a), 155.6*◦* (b), 158.2*◦* (c), 157.4*◦* (d)], H–N(6a–d) and C(2a– d)  $=$  O(2a–d) [N(6a–d)  $\cdots$  O(2a–d) = 2.99 Å (a), 3.09 Å (b), 2.96 Å (c), 3.10 A˚ (d); N–H ◊◊◊ O 160.8*◦* (a), 154.7*◦* (b), 160.2*◦* (c), 161.3*◦* (d)], H–N(8a–d) and C(4a–d)= $O(4a-d)$  [N(8a–d) $\cdots$ O(4a–d) = 3.00 Å (a), 2.96 Å (b), 3.00 Å (c), 2.93 Å (d); N–H ⋅ ⋅ · O 157.3<sup>°</sup> (a), 154.6*◦* (b), 157.1*◦* (c), 157.4*◦* (d)], and H–N(9a–d) and C(5a– d)  $=$  O(5a–d) [N(9a–d)  $\cdots$  O(5a–d) = 2.85 Å (a), 2.94 Å (b), 2.87 Å (c), 2.95 A˚ (d); N–H ◊◊◊ O 152.5*◦* (a), 168.9*◦* (b), 147.4*◦* (c), 162.8*◦* (d)]. Furthermore, one hydrogen bond of  $i \leftarrow i+3$  type between H–N(7a–d) and C(4a–d)= $O(4a-d)$  [N(7a–d)  $\cdots$  O(4a–d) = 2.98 Å (a), 3.02 A˚ (b), 3.06 A˚ (c), 3.04 A˚ (d); N–H ◊◊◊ O 128.2*◦* (a), 131.7*◦* (b), 150.1*◦* (c), 130.8*◦* (d)] was observed. In the packing mode of molecule **A**, four intermolecular hydrogen bonds were observed between the H–N(2a) donor and  $O_w(a)$  of the water (a) acceptor  $[N(2a)\cdots O_w(a) = 3.00 \text{ Å}; N-H \cdots O 124.1^{\circ}], H-N(3a) \text{ and } O_w(a)$  $[N(3a)\cdots O_w(a) = 3.00 \text{ Å}; N-H\cdots O 167.9^\circ]$ , the H–N(1a) and  $C(8a') = O(8a')$  [N(1a)  $\cdots$  O(8a') = 2.87 Å; N–H $\cdots$ O 162.0<sup>°</sup>] of a symmetry-related molecule  $(-1+x, y, z)$ , and the  $H-O_w(a)$  and C(7a′)=O(7a′)  $[O_w(a) \cdots O(7a') = 2.89 \text{ Å}; O-H \cdots O 176.6^{\circ}]$  of a symmetry-related molecule  $(-1+x, y, z)$ . For molecule **B**, six intermolecular hydrogen bonds were observed between  $H-O_w(b)$ and C(6b)=O(6b)  $[O_w(b) \cdots O(6b) = 2.91 \text{ Å}; O-H \cdots O \cdot 173.0^{\circ}],$ H–O<sub>w</sub>(b) and O<sub>w</sub>(c)  $[O_w(b) \cdots O_w(c) = 2.77 \text{ Å}; O-H \cdots O \frac{172.6^{\circ}}{172.6^{\circ}}]$ , H–O<sub>w</sub>(c) and C(7b)=O(7b)  $[O_w(c) \cdots O(7b) = 2.83$  Å; O–H $\cdots$ O 171.3<sup>°</sup>], the H–N(1b) and C(8b')= $O(8b')$  [N(1b) $\cdots$ O(8b') = 2.85 Å; N–H $\cdots$ O 167.0<sup>°</sup>] of a symmetry-related molecule (-1+x, y, z), the H–N(2b) and  $O_w(c') [N(2b) \cdots O_w(c') = 3.05 \text{ Å}; N-H \cdots \text{O}$ 125.5*◦*] of a symmetry-related molecule (-1+x, y, z), and the H– N(3b) and  $O_w(c')$  [N(3b) $\cdots$   $O_w(c') = 3.05$  Å; N–H $\cdots$  O 169.9<sup>°</sup>] of a symmetry-related molecule  $(-1+x, y, z)$ . For molecule **C**, four intermolecular hydrogen bonds were observed between  $H-O_w(d)$ and C(7c)=O(7c)  $[O_w(d) \cdots O(7c) = 2.90 \text{ Å}; O-H \cdots O 170.8^{\circ}],$ the H–N(1c) and C(8c')= $O(8c')$  [N(1c) $\cdots$ O(8c') = 2.85 Å; N–  $H \cdots$  O 159.3°] of a symmetry-related molecule (1+x, y, z), the H– N(2c) and O<sub>w</sub>(d') [N(2c)  $\cdots$  O<sub>w</sub>(d') = 2.99 Å; N–H  $\cdots$  O 128.7<sup>°</sup>] of a symmetry-related molecule  $(1+x, y, z)$ , and the H–N(3c) and  $O_w(d')$  $[N(3c)\cdots O_w(d') = 3.13$  Å (a bit long for an intermolecular hydrogen bond); N–H ··· O 167.1<sup>°</sup>] of a symmetry-related molecule (1+x, y, z). For molecule **D**, six intermolecular hydrogen bonds were observed between H–N(2d) and  $O_w(e)$  [N(2d) $\cdots$ O<sub>w</sub>(e) = 3.01 Å; N–H $\cdots$  O 129.6<sup>°</sup>], H–N(3d) and O<sub>w</sub>(e) [N(3d) $\cdots$ O<sub>w</sub>(e) = 3.17 Å (a bit long for an intermolecular hydrogen bond); N–H $\cdots$ O 168.3<sup>°</sup>, H–O<sub>w</sub>(f) and O<sub>w</sub>(e)  $[O_w(f) \cdots O_w(e) = 2.72 \text{ Å}; O-H \cdots \text{O}$ 170.8<sup>°</sup>], the H–N(1d) and C(8d')= $O(8d')$  [N(1d) $\cdots$ O(8d') = 2.81 Å; N–H $\cdots$  O 166.5<sup>°</sup>] of a symmetry-related molecule (1+x, y,

z), the H–O<sub>w</sub>(e) and C(7d')= $O(7d')$  [O<sub>w</sub>(e)  $\cdots$  O(7d') = 2.89 Å; O– H ⋅ ⋅ ⋅ O 177.2°] of a symmetry-related molecule (1+x, y, z), and the H–O<sub>w</sub>(f) and C(6d')= $O(6d')$  [O<sub>w</sub>(f)  $\cdots$  O(6d') = 2.89 Å; O–H $\cdots$  O 175.5*◦*] of a symmetry-related molecule (1+x, y, z). The four helical molecules **A–D** were connected by water molecule-mediated intermolecular hydrogen bonds, forming a head-to-tail alignment of chains ◊◊◊ **A** ◊◊◊ **A** ◊◊◊ **A** ◊◊◊ **A** ◊◊◊ , ◊◊◊**B**◊◊◊ **B**◊◊◊ **B**◊◊◊**B** ◊◊◊ , ◊◊◊ **C** ◊◊◊ **C** ◊◊◊ **C** ◊◊◊ **C** ◊◊◊ and ◊◊◊ **D** ◊◊◊ **D** ◊◊◊ **D** ◊◊◊ **D** ◊◊◊ , respectively (Fig. 11).



**Fig. 11** Packing of **6** (molecule **A**: blue, molecule **B**: purple, molecule **C**: green, molecule **D**: yellow) in the crystalline state. Intermolecular hydrogen bonds are indicated as red dashed lines.

In the asymmetric unit of Boc-[L-Leu-L-Leu- $(R,R)$ -Ac<sub>5</sub>c<sup>dOM</sup>]<sub>3</sub>-OMe  $(8)$ , two right-handed  $(P)$   $\alpha$ -helices were present with methanol and water molecules (Fig. 12). The conformations of the two molecules **A** and **B** were generally similar (Fig. 13), except for small differences at the N-terminus and at the side chains of the Leu and  $Ac_5c^{dOM}$  residues, and for the pattern of intramolecular hydrogen bonds. The mean values of the  $\phi$ and  $\psi$  torsion angles of the amino acid residues  $(1-8)$  were -66.3*◦* and -40.8*◦* for **A**, -59.4*◦* and -39.8*◦* for **B**, and reversal of the torsion angle signs at the C-terminus occurred at the Ac<sub>5</sub>c<sup>dOM</sup>(9) residue ( $\phi, \psi$  = +55.4<sup>°</sup>, +40.6<sup>°</sup> for **A** and



**Fig. 12** X-Ray diffraction structure of Boc-[L-Leu-L-Leu-  $(R, R)$ -Ac<sub>5</sub>c<sup>dOM</sup>]<sub>3</sub>-OMe (8), as viewed (A) perpendicular to and (B) along the helical axis. The methanol and water molecules are omitted.



**Fig. 13** The overlaid structures of molecules **A** (blue) and **B** (green), as viewed (A) perpendicular to and (B) along the helical axis.

 $\phi, \psi = +51.3^\circ$ , +46.4<sup>°</sup> for **B**). In molecule **A**, five intramolecular hydrogen bonds of the  $i \leftarrow i+4$  type between H–N(4a) and  $C(0a) = O(0a)$  [N(4a)  $\cdots$  O(0a) = 3.16 Å; N–H $\cdots$  O 166.0<sup>°</sup>], H– N(5a) and C(1a)=O(1a)  $[N(5a)\cdots O(1a) = 2.87 \text{ Å}; N-H\cdots O$ 156.5<sup>°</sup>], H–N(6a) and C(2a) = O(2a) [N(6a) $\cdots$ O(2a) = 2.93 Å;  $N-H \cdots O 166.6^{\circ}$ ],  $H-N(8a)$  and  $C(4a)=O(4a)$  [N(8a)  $\cdots O(4a)$  = 2.94 Å; N–H $\cdots$ O 147.4<sup>°</sup>], and H–N(9a) and C(5a)=O(5a)  $[N(9a)\cdots O(5a) = 2.82 \text{ Å}; N-H\cdots O(149.7^{\circ}]$  were observed. Furthermore, one weak intramolecular hydrogen bond was observed between H–N(7a) and C(3a)= $O(3a)$  [N(7a) $\cdots$ O(3a) = 3.27 Å; N–H $\cdots$ O 156.2<sup>°</sup>]. In the packing mode, six intermolecular hydrogen bonds between  $H-O_w(a)$  and  $C(6a) = O(6a)$  $[O_w(a) \cdots O(6a) = 2.87 \text{ Å}; O-H \cdots O(173.7^{\circ}], H-O_w(b)$  and C(9a)=O(9a)  $[O_w(b) \cdots O(9a) = 2.79 \text{ Å}; O-H \cdots O 174.0^\circ]$ , H–  $O_w(a)$  and  $O_w(c)$   $[O_w(a) \cdots O_w(c) = 2.82 \text{ Å}; O-H \cdots O 179.1^\circ]$ , H– N(1a) and C(8a')= $O(8a')$  [N(1a) $\cdots$  O(8a') = 2.83 Å; N–H $\cdots$ O 149.1*◦*] of a symmetry-related molecule (x, y, 1+z), the H–N(2a) and  $O_w(c')$  [N(2a) $\cdots$  $O_w(c') = 2.94$  Å; N–H $\cdots$ O 129.4<sup>°</sup>] of a symmetry-related molecule (x, y, 1+z), the H–O<sub>w</sub>(b) and O<sub>w</sub>(a')  $[O_w(b) \cdots O_w(a') = 3.02 \text{ Å}; O-H \cdots O 169.8^\circ]$  of a symmetry-related molecule (1+x, y, z), and one weak intermolecular hydrogen bond between the H–N(3a) and  $O_w(c')$  [N(3a) $\cdots$  $O_w(c') = 3.20$  Å; N–  $H \cdots$  O 169.8°] of a symmetry-related molecule (x, y, 1+z) were observed. In contrast to the *i*←*i*+4-type hydrogen bonds of molecule **A**, in molecule **B**, one intramolecular hydrogen bond of the  $i \leftarrow i+3$ type between H–N(3b) and  $C(0b) = O(0b)$   $[N(3b) \cdots O(0a) =$ 3.01 Å; N–H $\cdots$ O 149.2<sup>°</sup>] was observed. In addition, four intramolecular hydrogen bonds of the  $i \leftarrow i+4$  type between H–N(5b) and C(1b)=O(1b)  $[N(5b)\cdots O(1b) = 2.89 \text{ Å}; N-H\cdots O 166.0^{\circ}],$ H–N(6b) and C(2b)=O(2b) [N(6b)  $\cdots$  O(2b) = 2.92 Å; N–H $\cdots$  O 160.3<sup>°</sup>], H–N(8b) and C(4b)=O(4b) [N(8b) $\cdots$ O(4b) = 2.85 Å;  $N-H \cdots O 162.2^{\circ}$ ], H–N(9b) and C(5b)=O(5b) [N(9b) $\cdots$ O(5b) = 2.91 Å; N–H $\cdots$ O 157.7<sup>°</sup>] were observed. The distance between H-N(7b) and  $C(3b) = O(3b)$  (3.40 Å) was too long for an intramolecular hydrogen bond. In the packing mode, six intermolecular hydrogen bonds between  $H-O_w(d)$  and  $C(6b) = O(6b)$  $[O_w(d) \cdots O(6b) = 3.06 \text{ Å}; O-H \cdots O(172.1^{\circ})$ , the H– $O_w(e)$  and  $O_w(d)$   $[O_w(e) \cdots O_w(d) = 2.64$  Å;  $O-H \cdots O$  175.6<sup>°</sup>], the H– $O_w(e)$ and O<sub>MeOH</sub> of the methanol  $[O_w(e) \cdots O_{MeOH} = 2.87 \text{ Å}; O-H \cdots O$ 175.4<sup>°</sup>], the H–O<sub>MeOH</sub> and O<sub>M9</sub> of the methoxy group of  $(R, R)$ - $Ac_5c^{dOM}$  [O<sub>MeOH</sub>  $\cdots$  O<sub>M9</sub> = 2.77 Å; O–H $\cdots$  O 173.6<sup>°</sup>], the H–N(1b) and C(7b<sup>'</sup>)=O(7b') [N(1b)  $\cdots$  O(7b') = 2.82 Å; N–H $\cdots$ O 164.4<sup>°</sup>] of a symmetry-related molecule  $(x, y, 1+z)$ , and  $N(2b)$  and  $O_w(d')$  $[N(2b)\cdots O_w(d') = 2.89 \text{ Å}; N-H\cdots O 162.7^{\circ}]$  of a symmetryrelated molecule  $(x, y, 1+z)$  were observed. The  $\alpha$ -helical chains of **A** and **B** were packed forming a head-to-tail alignment of chains ◊◊◊ **A** ◊◊◊ **A** ◊◊◊ **A** ◊◊◊ **A** ◊◊◊ and ◊◊◊**B** ◊◊◊**B**◊◊◊ **B**◊◊◊ **B**◊◊◊ (Fig. 14).

The dominant conformations of four hexapeptides Cbz-(L-Leu- $L$ -Leu-dAA)<sub>2</sub>-OMe [dAA = 1: Aib; 2: Ac<sub>5</sub>c; 3: (*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup>; 4:  $(R, R)$ -Ac<sub>5</sub>c<sup>dOM</sup>] and four nonapeptides Boc-(L-Leu-L-Leu-dAA)<sub>3</sub>-OMe  $[dAA = 5$ : Aib; 6: Ac<sub>5</sub>c; 7: (*S*,*S*)-Ac<sub>5</sub>c<sup>dOM</sup>; 8: (*R*,*R*)-Ac<sub>5</sub>c<sup>dOM</sup>] in solution were found to be helical structures. Judging from the *R* value of the CD spectra in  $TFE/H<sub>2</sub>O$  solution and in the solid state, Aib hexapeptide 1 and Ac<sub>5</sub>c hexapeptide 2 formed righthanded ( $P$ ) 3<sub>10</sub>-helices, while Ac<sub>5</sub>c<sup>dOM</sup> hexapeptides 3 and 4 formed a mixture of (*P*)  $3_{10}$ - and  $\alpha$ -helices in solution and (*P*)  $\alpha$ -helices in the solid state. The CD spectra of Aib nonapeptide **5** suggested the existence of a  $(P)$  3<sub>10</sub>-helix in solution and in the solid state, while those of Ac<sub>5</sub>c nonapeptide 6 suggested the existence of a mixture of



**Fig. 14** Packing of **8** (molecule **A**: blue, molecule **B**: green) in the crystalline state. Intermolecular hydrogen bonds are indicated as red dashed lines.

(*P*)  $3_{10}$ - and  $\alpha$ -helices in solution and the existence of a  $3_{10}$ -helix in the solid state. On the other hand, the CD spectra of chiral  $Ac_5c^{dOM}$ nonapeptides 7 and 8 indicated the existence of  $(P)$   $\alpha$ -helices in solution and in the solid state. X-ray crystallographic analysis revealed that the Aib hexapeptide 1 assumed a  $(P)$  3<sub>10</sub>-helix, while  $(S, S)$ -Ac<sub>5</sub>c<sup>dOM</sup> hexapeptide 3 assumed a (*P*)  $\alpha$ -helix. Furthermore, the Ac<sub>5</sub>c nonapeptide 6 and  $(R,R)$ -Ac<sub>5</sub>c<sup>dOM</sup> nonapeptide 8 were folded into  $(P)$   $\alpha$ -helices in the crystal state, and there was not much difference with the packing of the five-membered ring of the Ac<sub>5</sub>c and Ac<sub>5</sub>c<sup>dOM</sup> residues (Fig. 15).



**Fig. 15** The overlaid structures of nonapeptides **6** (molecule **A**: blue) and **8** (molecule **A**: green), as viewed (A) perpendicular to and (B) along to the helical axis.

Aib and  $Ac<sub>5</sub>c$  are achiral amino acids, and thus do not have the bias for the helical-screw handedness. Therefore, the righthanded helical screw senses of L-Leu-based hexapeptides **1** and **2** and nonapeptides **5** and **6** were controlled by the chiral centers of L-Leu  $\alpha$ -carbon atoms. On the other hand,  $(S, S)$ -Ac<sub>5</sub>c<sup>dOM</sup> and  $(R,R)$ -Ac<sub>5</sub>c<sup>dOM</sup> are chiral cyclic  $\alpha$ , $\alpha$ -disubstituted  $\alpha$ -amino acids, having chiral centers only at the side chain. The relationship between  $(S, S)$  and  $(R, R)$  is enantiomeric, thus the helical-screw properties of these cyclic amino acids should be opposite. All L-Leu hexapeptides and nonapeptides having chiral  $Ac_5c^{dOM}$  showed right-handed helical-screw sense. These results mean that L-Leu chiral centers on the  $\alpha$ -carbon atoms are stronger than those of the side-chain chiral centers of  $Ac_5c^{dOM}$ . The Aib and achiral  $Ac_5c$ residues have the tendency to form  $(P)$  3<sub>10</sub>-helices in short L-Leu peptides, whereas the chiral  $Ac_5c^{dOM}$  residues have a penchant for forming  $(P)$   $\alpha$ -helices in short L-Leu peptides. The  $\alpha$ -helical property of chiral  $Ac_5c^{dOM}$  may be due to the influence of the methoxy substituents, especially in terms of hydrophilicity and bulkiness on the cyclopentane ring. Therefore, the introduction of chiral  $Ac_5c^{dOM}$  residues at *i* and  $i+3$  positions in the peptide may stabilize not a  $3_{10}$ -helix but an  $\alpha$ -helix by avoiding contact with the methoxy substituents of  $Ac_5c^{dOM}$  at *i* and *i*+3 positions.

## **Conclusions**

In conclusion, four types of  $\alpha, \alpha$ -disubstituted amino acids: Aib, Ac<sub>5</sub>c,  $(S, S)$ -Ac<sub>5</sub>c<sup>dOM</sup> and  $(R, R)$ -Ac<sub>5</sub>c<sup>dOM</sup>, were introduced into the L-leucine-based peptides and their influence on peptide conformation was studied. We demonstrated that the well-known achiral  $\alpha$ , $\alpha$ -disubstituted amino acid Aib induces the 3<sub>10</sub>-helical structure in short peptides, but the newly designed chiral cyclic  $\alpha$ , $\alpha$ -disubstituted amino acids Ac<sub>5</sub>c<sup>dOM</sup> preferentially induce the  $\alpha$ helical structure. Although it has been believed that at least seven amino acid residues are necessary to construct  $\alpha$ -helical peptides in the crystal state,**3,35–38** the L-Leu-hexapeptides containing chiral  $Ac_5c^{dOM}$  formed  $\alpha$ -helices.<sup>24,39</sup> These results provide valuable information for the design of stable helical peptides and may also be applicable in many fields such as organic, bioorganic and medicinal chemistries.

## **Experimental**

## **General remarks**

FT-IR spectra were recorded on a Nicolet Avatar-320 spectrometer at 1 cm-<sup>1</sup> resolution, with an average of 128 scans used for the solution  $(CDCl_3)$  method and a 0.1 mm path length for NaCl cells, or used for the attenuated total reflection (ATR) method.

CD spectra were recorded with a *Jasco J-720W* spectropolarimeter using a 1.0 mm path length cell. The data are expressed in terms of  $[\theta]_M$ , the total molar ellipticity (deg cm<sup>2</sup> dmol<sup>-1</sup>). A mixture of 2,2,2-trifluoroethanol (TFE) and  $H<sub>2</sub>O (50/50)$  was used as a solvent in solution, and a KCl disk was used in the solid state.

Single crystals of peptides **1**, **3** and **8** were grown from MeOH/H<sub>2</sub>O, and that of 6 was from EtOH/H<sub>2</sub>O. Data collection was performed on a Bruker AXS SMART 1000 CCD imaging plate diffractometer using graphite-monochromated Mo- $K\alpha$  radiation. All crystals remained stable during the X-raydata collection. The structures of the crystals were solved using the SHELXS 97**<sup>40</sup>** direct method and expanded by the Fourier technique.**<sup>41</sup>** All non-H-atoms were given anisotropic thermal parameters, some H-atoms were refined isotropically, and the remaining H-atoms at the calculated positions were given isotropic thermal parameters. The final cycle of full-matrix least-squares refinement of 1 gave an  $R_1$  factor of 0.0434 based on 3417 ( $I >$  $2\sigma(I)$ ) reflections and an *Rw* factor of 0.0870 for all data. The *R<sub>1</sub>* factor of **3** was 0.0522 based on 12192 ( $I > 2\sigma(I)$ ) reflections and an *Rw* factor of 0.1427 for all data. The *R1* factor of **6** was 0.0962 based on 21523 ( $I > 2\sigma(I)$ ) reflections and an *Rw* factor of 0.2635 for all data. The  $R_1$  factor of 8 was 0.0859 based on 15919 ( $I >$  $2\sigma(I)$ ) reflections and an *Rw* factor of 0.2370 for all data. All data for peptides **1**, **3**, **6** and **8** have been deposited in the Cambridge Crystallographic Data Centre (CCDC).†

#### Cbz-(L-Leu-L-Leu-Aib)<sub>2</sub>-OMe (1)

Colorless crystals; mp 195–197 <sup>°</sup>C (recryst. from MeOH); [α]<sup>25</sup> -11.4 (*c* 1.3, CHCl<sub>3</sub>); IR (ATR solid state) 3314, 2956, 1745, 1654, 1527 cm-<sup>1</sup> ; 1 H NMR (400 MHz, CDCl3) *d* 7.55 (d, *J* = 7.8 Hz, 1H), 7.31–7.40 (m, 5H), 7.23–7.24 (br s, 2H), 7.19 (br s, 1H), 7.01 (br s, 1H), 6.32 (br s, 1H), 5.15 (dd, *J* = 12.4, 39.0 Hz, 2H), 4.33 (m, 1H), 4.17 (m, 1H), 3.87–3.98 (m, 2H), 3.65 (s, 3H), 1.40–1.87 (m, 24H), 0.73–0.97 (m, 24H); FAB(+)-MS  $m/z$  811.4 (M<sup>+</sup>+Na); elemental analysis calcd for  $C_{41}H_{68}O_9N_6$ : C 62.41, H 8.69, N 10.65: found C 62.49, H 8.57, N 10.78.

## Cbz-(L-Leu-L-Leu-Ac<sub>5</sub>c)<sub>2</sub>-OMe (2)

Colorless crystals; mp 98–100  $\rm{°C}$  (recryst. from CHCl<sub>3</sub>); [ $\alpha$ ]<sup>25</sup> –1.5 (*c* 1.0, CHCl3); IR (ATR solid state) 3322, 2957, 1744, 1652, 1525 cm-<sup>1</sup> ; 1 H NMR (400 MHz, CDCl3) *d* 7.56 (d, *J* = 9.2 Hz, 1H), 7.42 (br s, 1H), 7.34–7.37 (m, 5H), 7.25 (br s, 1H), 7.17 (br s, 1H), 6.79 (br s, 1H), 5.68 (br s, 1H), 5.15 (dd, *J* = 12.4, 28.4 Hz, 2H), 4.10–4.16 (m, 2H), 3.88–3.95 (m, 2H), 3.69 (s,3H), 2.56 (m, 1H), 2.00–2.25 (m, 5H), 1.50–1.90 (m, 22H), 0.87–0.98 (m, 24H); FAB(+)-MS  $m/z$  863.5 (M<sup>+</sup>+Na).

# $Cbz$ -[L-Leu-L-Leu- $(S<sub>1</sub>,S)$ -Ac<sub>5</sub>c<sup>dOM</sup>]<sub>2</sub>-OMe (3)

Colorless crystals; mp 115–117 °C (recryst. from MeOH);  $[\alpha]_D^{24}$  = +0.7 (*c* = 0.6, CHCl3); IR (ATR solid state) 3335, 2956, 1742, 1654, 1531 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.54 (br s, 1H), 7.32–7.45 (m, 8H), 7.15 (br s, 1H), 6.24 (br s, 1H), 5.18 (dd, *J* = 12.4, 34.1 Hz, 2H), 4.32 (m, 1H), 4.21 (m, 1H), 3.67–4.01 (m, 6H), 3.64 (s, 3H), 3.31–3.35 (m, 12H), 2.94 (dd, *J* = 7.3, 14.0 Hz, 1H), 2.55–2.96 (m, 2H), 2.39–2.44 (m, 2H), 2.28 (dd, *J* = 7.3, 14.2 Hz, 1H), 2.07–2.17 (m, 2H), 1.97 (dd, *J* = 7.3, 13.9 Hz, 1H), 1.50–1.87 (m, 11H), 0.77–1.08 (m, 24H); FAB(+)-MS *m*/*z* 983.6 (M++Na); elemental analysis calcd for  $C_{49}H_{80}O_{13}N_6$ : C 61.23, H 8.39, N 8.74: found C 61.24, H 8.35, N 8.98.

## $C$ bz-[L-Leu-L-Leu- $(R,R)$ -Ac<sub>5</sub>c<sup>dOM</sup>]<sub>2</sub>-OMe (4)

Colorless crystals; mp 118–120 °C (recryst. from MeOH); [α]<sup>22</sup> -22.9 (*c* 0.8, CHCl3); IR (ATR solid state) 3321, 2957, 1743, 1652, 1534 cm-<sup>1</sup> ; 1 H NMR (400 MHz, CDCl3) *d* 7.52 (br s, 1H), 7.43 (br s, 1H), 7.34–7.39 (m, 7H), 7.00 (br s, 1H), 6.30 (br s, 1H), 5.20 (s, 2H), 4.34 (m, 1H), 4.22 (m, 1H), 3.73–3.95 (m, 6H), 3.62 (s, 3H), 3.29–3.33 (m, 12H), 3.05 (dd, *J* = 7.5, 14.4 Hz, 1H), 2.83 (dd, *J* = 7.1, 13.9 Hz, 1H), 2.58–2.66 (m, 2H), 2.28 (d, *J* = 19.3 Hz, 1H), 2.24 (dd, *J* = 7.1, 14.2 Hz, 1H), 1.49–2.02 (m, 14H), 0.83–0.99 (m, 24H): FAB(+)-MS  $m/z$  983.6 (M<sup>+</sup>+Na); elemental analysis calcd for  $C_{49}H_{80}O_{13}N_6$ : C 61.23, H 8.39, N 8.74: found C 61.58, H 8.43, N 8.79.

## Boc-(L-Leu-L-Leu-Aib)<sub>3</sub>-OMe (5)

Colorless crystals; mp 191–193 °C (recryst. from MeOH); [α]<sup>24</sup><sub>D</sub> -15.1 (*c* 1.0, CHCl3); IR (ATR solid state) 3315, 2961, 1734, 1658, 1531 cm-<sup>1</sup> ; 1 H NMR (400 MHz, CDCl3) *d* 7.26–7.48 (m, 7H), 6.83 (d, *J* = 4.1 Hz, 1H), 5.31 (d, *J* = 2.8 Hz, 1H), 4.32 (m, 1H), 4.14 (m, 1H), 3.85–4.04 (m, 4H), 3.66 (s, 3H), 1.40–1.79 (m, 45H), 0.84–1.09 (m, 36H); FAB(+)-MS  $m/z$  1088.7 (M<sup>+</sup>+Na); elemental analysis calcd for  $C_{54}H_{99}O_{12}N_9$ : C 60.82, H 9.36, N 11.82: found C 60.73, H 9.34, N 11.91.

#### $Boc$ -(L-Leu-L-Leu-Ac<sub>5</sub>c)<sub>3</sub>-OMe (6)

Colorless crystals; mp 213–215 °C (recryst. from CHCl<sub>3</sub>);  $[\alpha]_D^{24} =$ +6.8 (*c* = 1.0, CHCl3); IR (ATR solid state) 3313, 2957, 1740, 1652, 1529 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.66 (d, *J* = 4.6 Hz, 1H), 7.57 (br s, 1H), 7.45–7.48 (m, 3H), 7.35 (d, *J* = 5.9 Hz, 1H), 7.24 (br s, 1H), 6.55 (br s, 1H), 5.06 (br s, 1H), 4.34 (m, 1H), 4.19 (m, 1H), 3.88–4.02 (m, 4H), 3.66 (s, 3H), 2.60–2.65 (m, 2H), 2.07–2.24 (m, 6H), 1.58–1.94 (m, 34H), 1.56 (s, 9H), 0.79–1.00 (m, 36H): FAB(+)-MS *m*/*z* 1144 (M++H); elemental analysis calcd for  $C_{60}H_{105}O_{12}N_9$ : C 62.96, H 9.25, N 11.01: found C 62.94, H 9.24, N 10.86.

#### $Boc$ -[L-Leu-L-Leu- $(S, S)$ -Ac<sub>5</sub>c<sup>dOM</sup>]<sub>3</sub>-OMe (7)

Colorless crystals; mp 110–112 °C (recryst. from MeOH);  $[\alpha]_D^{24}$  =  $+19.6$  ( $c = 1.35$ , CHCl<sub>3</sub>); IR (ATR solid state); 3321, 2960, 1736, 1661, 1529 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.68 (d, *J* = 4.6 Hz, 1H), 7.34–7.46 (m, 6H), 6.94 (d, *J* = 2.6 Hz, 1H), 5.22 (d, *J* = 3.7 Hz, 1H), 4.28 (m, 1H), 4.18 (m, 1H), 3.89–4.05 (m, 5H), 3.67–3.83 (m, 5H), 3.66 (s, 3H), 3.30–3.36 (m, 18H), 2.97 (dd, *J* = 7.3, 13.8 Hz, 1H), 2.63–2.77 (m, 3H), 2.18–2.37 (m, 5H), 2.08 (dd, *J* = 7.6, 14.2 Hz, 1H), 1.52–2.03 (m, 20H), 1.49 (s, 9H), 0.88–1.10 (m, 36H); FAB(+)-MS  $m/z$  1346.9 (M<sup>+</sup>+Na); elemental analysis calcd for  $C_{66}H_{117}O_{18}N_9$ : C 59.84, H 8.90, N 9.52: found C 60.17, H 8.91, N 9.65.

#### $Boc$ -[L-Leu-L-Leu- $(R,R)$ -Ac<sub>5</sub> $c^{dOM}$ ]<sub>3</sub>-OMe (8)

Colorless crystals; mp 170–172 °C (recryst. from MeOH); [α]<sup>24</sup><sub>D</sub> -6.65 (*c* 0.85, CHCl<sub>3</sub>); IR (ATR solid state) 3330, 2960, 1736, 1658, 1532 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.75 (d, *J* = 5.1 Hz, 1H), 7.70 (br s, 1H), 7.58 (d, *J* = 5.3 Hz, 1H), 7.55 (br s, 1H), 7.51 (br s, 1H), 7.40 (d, *J* = 5.05 Hz, 1H), 7.38 (br s, 1H), 6.72 (br s, 1H), 5.43 (br s, 1H), 4.36 (m, 1H), 4.20 (m, 1H), 3.76– 4.00 (m, 10H), 3.69 (s, 3H), 3.25–3.38 (m, 18H), 3.19 (dd, *J* = 7.6, 14.3 Hz, 1H), 3.01 (dd, *J* = 6.2, 14.4 Hz, 1H), 2.70–2.94 (m, 2H), 2.64–2.68 (m, 2H), 2.14–2.24 (m, 2H), 1.61–2.00 (m, 22H), 1.54 (s, 9H), 0.85–1.02 (m, 36H); FAB(+)-MS *m*/*z* 1346.9 (M++Na); elemental analysis calcd for  $C_{66}H_{117}O_{18}N_9$ : C 59.84, H 8.90, N 9.52: found C 60.15, H 8.91, N 9.62.

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#### **Notes and references**

- 1 J. P. Schneider and J. W. Kelly, *Chem. Rev.*, 1995, **95**, 2169– 2187.
- 2 J. Venkatraman, S. C. Shankaramma and P. Balaram, *Chem. Rev.*, 2001, **101**, 3131–3152.
- 3 C. Toniolo, M. Crisma, F. Formaggio, C. Peggion, Q. B. Broxterman and B. Kaptein, *J. Inclusion Phenom. Macrocyclic Chem.*, 2005, **51**, 121–136.
- 4 S. H. Gellman, *Acc. Chem. Res.*, 1998, **31**, 173–180.
- 5 H. Heimgartner, *Angew. Chem., Int. Ed. Engl.*, 1991, **30**, 238–264.
- 6 E. Benedetti, V. Barone, A. Bavoso, D. B. Blasio, F. Lelj, V. Pavone, C. Pedone, G. M. Bonora, C. Toniolo, M. T. Leplawy, K. Kaczmarek and A. Redlinski, *Biopolymers*, 1988, **27**, 357–371.
- 7 C. Toniolo, G. M. Bonora, A. Bavoso, E. Benedetti, B. Di Blasio, V. Pavone, C. Pedone, V. Barone, F. Lelj, M. T. Leplawy, K. Kaczmarek and A. Redlinski, *Biopolymers*, 1988, **27**, 373–379.
- 8 C. Toniolo, M. Crisma, F. Formaggio, C. Valle, G. Cavicchioni, G. Precigoux, A. Aubry and J. Kamphuis, *Biopolymers*, 1993, **33**, 1061– 1072.
- 9 F. Formaggio, A. Barazza, A. Bertocco, C. Toniolo, Q. B. Broxterman, B. Kaptein, E. Brasola, P. Pengo, L. Pasquato and P. Scrimin, *J. Org. Chem.*, 2004, **69**, 3849–3856.
- 10 M. Crisma, M. Saviano, A. Moretto, Q. B. Broxterman, B. Kaptein and C. Toniolo, *J. Am. Chem. Soc.*, 2007, **82**, 15471–15473.
- 11 B. Jaun, M. Tanaka, P. Seiler, F. N. M. Kuhnle, C. Braun and D. ¨ Seebach, *Liebigs Ann./Recl.*, 1997, 1697–1710.
- 12 M. Tanaka, *Chem. Pharm. Bull.*, 2007, **55**, 349–358.
- 13 I. L. Karle and P. Balaram, *Biochemistry*, 1990, **29**, 6747–6756.
- 14 M. Crisma, E. Andreetto, M. D. Zotti, A. Moretto, C. Peggion, F. Formaggio and C. Toniolo, *J. Pept. Sci.*, 2007, **13**, 190–205.
- 15 S. Aravinda, N. Shamala and P. Balaram, *Chem. Biodiversity*, 2008, **5**, 1238–1262.
- 16 A. Moretto, I. Menegazzo, M. Crisma, E. J. Shotton, H. Nowell, S. Mammi and C. Toniolo, *Angew. Chem., Int. Ed.*, 2009, **48**, 8986–8989.
- 17 M. Oba, Y. Demizu, N. Yamagata, Y. Sato, M. Doi, M. Tanaka, H. Suemune, H. Okuda and M. Kurihara, *Tetrahedron*, 2010, **66**, 2293– 2296.
- 18 Y. Demizu, N. Yamagata, Y. Sato, M. Doi, M. Tanaka, H. Okuda and M. Kurihara, *J. Pept. Sci.*, 2010, **16**, 153–158.
- 19 Y. Demizu, M. Doi, Y. Sato, M. Tanaka, H. Okuda and M. Kurihara, *J. Org. Chem.*, 2010, **75**, 5234–5239.
- 20 M. Tanaka, Y. Demizu, M. Doi, M. Kurihara and H. Suemune, *Angew. Chem., Int. Ed.*, 2004, **43**, 5360–5363.
- 21 Y. Demizu, M. Tanaka, M. Doi, M. Kurihara, H. Okuda and H. Suemune, *J. Pept. Sci.*, 2010, **16**, 621–626.
- 22 S. Royo, W. M. De Borggraeve, C. Peggion, F. Formaggio, M. Crisma, A. I. Jiménez, C. Cativiela and C. Toniolo, *J. Am. Chem. Soc.*, 2005, **127**, 2036–2037.
- 23 M. Tanaka, K. Anan, Y. Demizu, M. Kurihara, M. Doi and H. Suemune, *J. Am. Chem. Soc.*, 2005, **127**, 11570–11571.
- 24 Preliminary communication of this paper was published. See: Y. Demizu, M. Tanaka, M. Nagano, M. Kurihara, M. Doi, T. Maruyama and H. Suemune, *Chem. Pharm. Bull.*, 2007, **55**, 840–842.
- 25 M. Nagano, M. Doi, M. Kurihara, H. Suemune and M. Tanaka, *Org. Lett.*, 2010, **12**, 3564–3566.
- 26 J. Sheehan, P. Cruickshank and G. Boshart, *J. Org. Chem.*, 1961, **26**, 2525–2528.
- 27 J. Sheehan and G. P. Hess, *J. Am. Chem. Soc.*, 1955, **77**, 1067–1068.
- 28 D. F. Kennedy, M. Crisma, C. Toniolo and D. Chapman, *Biochemistry*, 1991, **30**, 6541–6548.
- 29 M. Tanaka, S. Nishimura, M. Oba, Y. Demizu, M. Kurihara and H. Suemune, *Chem.–Eur. J.*, 2003, **9**, 3082–3090.
- 30 C. Toniolo, G. M. Bonora, A. Bavoso, E. Benedetti, B. Di Blasio, V. Pavone, C. Pedone, V. Barone, F. Lelj, M. T. Leplawy, K. Kaczmarek and A. Redlinski, *Biopolymers*, 1988, **27**, 373–379.
- 31 C. Toniolo, A. Polese, F. Formaggio, M. Crisma and J. Kamphuis, *J. Am. Chem. Soc.*, 1996, **118**, 2744–2745.
- 32 F. Formaggio, M. Crisma, P. Rossi, P. Scrimin, B. Kaptein, Q. B. Broxterman, J. Kamphuis and C. Toniolo, *Chem.–Eur. J.*, 2000, **6**, 4498– 4504.
- 33 L. Pal, G. Basu and P. Chakrabarti, *Proteins: Struct., Funct., Genet.*, 2002, **48**, 571–579.
- 34 R. Bardi, A. M. Pizzesi, C. Toniolo, M. Sukumar and P. Balaram, *Biopolymers*, 1986, **25**, 1635–1644.
- 35 C. Toniolo, M. Crisma, F. Formaggio, C. Peggion, Q. B. Broxterman and B. Kaptein, *Biopolymers*, 2004, **76**, 162–176.
- 36 M. Crisma, F. Formaggio, A. Moretto and C. Toniolo, *Biopolymers*, 2006, **84**, 3–12.
- 37 V. Pavone, E. Benedetti, B. Di Blasio, C. Pedone, A. Santini, A. Bavoso, C. Toniolo, M. Crisma and L. Satore, *J. Biomol. Struct. Dyn.*, 1990, **7**, 1321–1331 .
- 38 M. Crisma, M. Saviano, A. Moretto, Q. B. Broxterman, B. Kaptein and C. Toniolo, *J. Am. Chem. Soc.*, 2007, **129**, 15471–15473.
- 39 Six residues are enough to construct an  $\alpha$ -helical peptide in solution: A. Moretto, F. Formaggio, B. Kaptein, Q. B. Broxterman, L. Wu, T. A. Keiderling and C. Toniolo, *Biopolymers*, 2008, **90**, 567–574.
- 40 G. M. Sheldrick, *SHELXL 97. Program for Crystal Structure Refinement.* University of Göttingen: Göttingen (1997).
- 41 P. T. Beurskens, G. Admiraal, G. Beurskens, W. P. Bosman, R. de Gelder, R. Israel, J. M. M. Smits, *The DIRDIF-99 program system, Technical Report of the Crystallography Laboratory*, University of Nijmegen, The Netherlands (1994).